

Immunodetection of N-Glycolyl GM3 Ganglioside in Formalin-Fixed and Paraffin-Embedded Tissues: A Fact that Needs Further Investigations

Rances Blanco

¹Laboratory of Recognition and Biological Activity Assays, Center of Molecular Immunology, Havana, Cuba.

***Corresponding author:** Rances Blanco, Center of Molecular Immunology, 216 St and 15 Ave, Atabey, Playa. PO Box 16040, Havana 11600, Cuba, Phone: +5372717933-3464, Fax: +5372720644, E-mail: rances@cim.sld.cu

Received Date: 14th April 2018

Accepted Date: 16th April 2018

Published Date: 20th April 2018

Citation: Blanco R (2018) Immunodetection of N-Glycolyl GM3 Ganglioside in Formalin-Fixed and Paraffin-Embedded Tissues: A Fact that Needs Further Investigations. Enliven:Immunol Immunotechnol 5(1):00e1

Copyright: © 2018 Rances Blanco. This is an Open Access article published and distributed under the terms of the Creative Commons Attribution License, that permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Editorial

Gangliosides are glycosphingolipids containing at least one residue of sialic acid [1]. These molecules are mostly localized on cell membrane with the sialylated sugar chains protruding out of cells and the ceramide moiety anchor in the outer surface of plasmatic membranes [2]. Traditionally, immunohistochemistry (IHC) has permitted the detection of a variety of gangliosides in normal and malignant human tissues [3]. Among these molecules, N-glycolyl GM3 ganglioside (NeuGcGM3) has been immunohistochemically localized in human tumors, becoming both a prognostic factor and an attractive target for cancer immunotherapy [4,5].

Frozen tissues are considered the most adequate kind of samples for the study of gangliosides. In line with this, an increased expression of NeuGcGM3 was demonstrated in some malignant tumors [5-7] using frozen tissues after 4% paraformaldehyde (methanol-free formaldehyde) fixation. Regrettably, the number of cases included in these studies was limited, due to frozen tissues are not available in sufficiently large quantities in conventional pathology departments. This fact allowed using formalin-fixed and paraffin-embedded (FFPE) samples which are the most common specimens available for NeuGcGM3 analysis after the histopathological diagnosis.

Based on the localization and the chemical composition of NeuGcGM3, at least three questions related with the immunodetection of this molecule in FFPE samples exist: (1) the organic solvents used in the routine tissue processing (e.g. methanol, ethanol) removes an important fraction of NeuGcGM3 from the cell surface, (2) the specificity of monoclonal antibodies (mAb) that usually recognize oligosaccharides chains associated to both glycosphingolipids and glycoproteins [8,9] and (3) the increased content of NeuGcGM3 detected in human tumors since gangliosides, with exception of brain, commonly constitute less than 5% of lipids present in cell membranes [10,11].

It is known that after formaldehyde fixation carbohydrates, lipids and nucleic acids are trapped in a matrix of insolubilized and cross-linked proteins. But, the chemical structure of these molecules is not altered by formaldehyde unless fixation is prolonged for several weeks [12]. Although, commercial formaldehyde solution contains about 10-15% of methanol to prevent the polymerization, it is insufficient to completely remove NeuGcGM3 from human tissues after fixation and subsequent tissue processing [4,13]. However, in cases of lower levels of expression, NeuGcGM3 could be totally extracted from tissue sections, affecting the selection of candidates for immunotherapy.

Alonso et al. reported a comparable reaction of GM3-immunized sera against both paraffin and resins embedding GM3-expressing melanoma tumors, suggesting that the conventional histological processing is incapable to extract and/or damage the antigenic carbohydrate determinants of gangliosides [14]. Tissue processing for electron microscopy using resins such as Epon minimizes the extraction of lipids. In a similar way, Carr et al. reported a strong reactivity against FFPE breast carcinoma sections using sera from selected patients immunized with the NGcGM3/VSSP vaccine. This is a cancer vaccine able to induce a specific immune response against NeuGcGM3 [15].

In addition, P3 mAb was able to recognize Wilm's tumors and non-small cell lung cancer (NSCLC) [6,16] in FFPE samples, while the reactivity of GMR8 Mab was evidenced in the later [13]. These mAb are IgMs that react to NeuGcGM3 and other NeuGc-containing gangliosides [17,18]. However, NeuGcGM3 was the most NeuGc-containing ganglioside detected in NSCLC [13]. Interestingly, Hayashi et al. demonstrated that chloroform-methanol was capable to completely remove the reactivity of GMR8 mAb from NSCLC sections. Furthermore, the conservation of

NeuGcGM3 and NeuGcGD1a in the lipidic fraction of FFPE samples was demonstrated by mean of TLC-immunostaining [13].

The staining of 14F7 mAb, a highly specific IgG1 against NeuGcGM3 [5], has been evidenced in a variety of FFPE human tumors [6,16,19]. This mAb is able to discriminate NeuGcGM3 from N-acetyl GM3 (a closely related molecule) [5,20], which is a normal constituent of human cells. Moreover, the specificity of 14F7 mAb reaction was confirmed by enzymatic treatment of FFPE sections [4], although a faintly decreased in the staining after protease exposure was observed. In this regard, it was suggested that 14F7 mAb reacts with the oligosaccharide core of NeuGcGM3 present in glycolipids and glycoproteins [5], potentially enriching the staining of this mAb.

A correlation between the in vivo radioimmunolocalization of NeuGcGM3-expressing breast tumors using ^{99m}Tc labeled 14F7 and the reactivity of this mAb in the FFPE counterparts by IHC was also reported [21]. Similarity, the immunostaining of 14F7 mAb in frozen tissues correlated with the FFPE counterparts [7]. Furthermore, it was demonstrated that treatment with ethanol and methanol was unable to extract NeuGcGM3 from NSCLC sections after formaldehyde fixation [7]. This molecule was also found in lipidic extracts obtained from FFPE samples by mean of TLC-immunostaining with 14F7 mAb and mass spectrometry [4].

Finally, an over expression of GM3 synthase gene usually occurs in malignant cells, inducing an augment in the NeuAcGM3 content [22-24]. In addition, the hypoxic condition of tumors provokes the over expression of sialin, a sialic acid transporter, which favors the absorption of NeuGc from the external medium and its posterior incorporation to newly synthesized glycoconjugates. Consequently, an increase in the amount of NeuGc-containing gangliosides, including NeuGcGM3, takes place in malignancies [24,25]. Moreover, a change from NeuAcGM3 to NeuGcGM3 could also occur in tumors, as it was previously described in animal models [26].

All these facts support the increased proportion of NeuGcGM3 found in FFPE tumor sections. They also support the use of FFPE tissues as a selection tool of cancer patient candidates for specific therapies using NeuGcGM3 as target. However, the molecular basis to understand the mechanism by which NeuGcGM3 is preserved in this kind of samples needs further investigations.

Acknowledgments

The author wants to express grateful to Ph.D. Adriana Carr (Center of Molecular Immunology) for the valuable discussion about the paper as well as to Mr. Rolando Dominguez for his editorial assistance.

References

1. Yu RK, Tsai YT, Aria T, Yanagisawa M (2011) Structures, biosynthesis, and functions of gangliosides-an overview. *J Oleo Sci* 60: 537-544.
2. Sonnino S, Prinetti A (2010) Gangliosides as regulators of cell membrane organization and functions. In: *Sphingolipids as Signaling and Regulatory Molecules* 688: 165-184.
3. Zhang S, Cordon-Cardo C, Zhang HS, Reuter VE, Adluri S, et al. (1997) Selection of tumor antigens as targets for immune attack using immunohistochemistry: I. Focus on gangliosides. *Int J Cancer* 73: 42-49.
4. Blanco R, Dominguez E, Morales O, Blanco D, Martinez D, et al. (2015) Prognostic significance of N-Glycolyl GM3 ganglioside expression in non-small cell lung carcinoma patients: new evidences. *Patholog Res Int* 2015: 132326.
5. Carr A, Mullet A, Mazorra Z, Vazquez AM, Alfonso M, et al. (2000) A mouse IgG1 monoclonal antibody specific for N-glycolyl GM3 ganglioside recognized breast and melanoma tumors. *Hybridoma* 19: 241-247.
6. Blanco R, Quintana Y, Blanco D, Cedeno M, Rengifo ChE, et al. (2013) Tissue reactivity of the 14F7 Mab raised against N-glycolyl GM3 ganglioside in tumors of neuroectodermal, mesodermal and epithelial origin. *J Biomark* 2013: 602417.
7. Blanco R, Rengifo ChE, Cedeno M, Frometa M, Hernandez T, et al. (2014) Immunodetection of N-glycolyl GM3 ganglioside in lung carcinoma by immunohistochemistry: a technical study using frozen and formalin-fixed and paraffin-embedded tissues. *Acta Microscopica* 23: 199-213.
8. Muthing J, Steuer H, Peter-Katalinic J, Marx U, Bethke U, et al. (1994) Expression of gangliosides GM3 (NeuAc) and GM3 (NeuGc) in myelomas and hybridomas of mouse, rat and human origin. *J Biochem* 116: 64-73.
9. Kovacic N, Muthing J, Marusic A (2000) Immunohistological and flow cytometric analysis of glycosphingolipid expression in mouse lymphoid tissues. *J Histochem Cytochem* 48: 1677-1690.
10. Singh SP, Tomar BS (2008) *Cell Biology* (9th edtn), Rastogi Publications, New Delhi, India.
11. Alberts B, Johnson A, Lewis J, Walter P, Raff M, et al. (2002) *The Lipid Bilayer*. In: *Molecular Biology of the Cell* (4th edition), Garland Science.
12. Kiernan JA (2000) Formaldehyde, formalin, paraformaldehyde and glutaraldehyde: What they are and what they do. *Microscopy Today* 1: 8-12.
13. Hayashi N, Chiba H, Kuronuma K, Go S, Hasegawa Y, et al. (2013) Detection of N-glycolylated gangliosides in non-small-cell lung cancer using GMR8 monoclonal antibody. *Cancer Sci* 104: 43-47.
14. Alonso DF, Gabri MR, Guthmann MD, Fainboim L, Gomez DE (1999) A novel hydrophobized GM3 ganglioside/Neisseria meningitidis outer membrane protein complex vaccine induces tumor protection in B16 murine melanoma. *Int J Oncol* 15: 59-66.
15. Carr A, Rodriguez E, Arango MC, Camacho R, Osorio M, et al. (2003) Immunotherapy of Advanced Breast Cancer With a Heterophilic Ganglioside (NeuGcGM3) Cancer Vaccine. *J Clin Oncol* 21: 1015-1021.
16. Scursioni AM, Galluzzo L, Camarero S, Pozzo N, Gabri MR, et al. (2010) Detection and characterization of N-glycolylated gangliosides in Wilms tumor by immunohistochemistry. *Pediatr Dev Pathol* 13: 18-23.
17. Vazquez AM, Alfonso M, Lanne B, Karlsson KA, Carr A, et al. (1995) Generation of a murine monoclonal antibody specific for N-glycolylneuraminic acid-containing gangliosides that also recognizes sulfated glycolipids. *Hybridoma* 14: 551-556.
18. Ozawa H, Kawashima I, Tai T (1992) Generation of murine monoclonal antibodies specific for N-glycolylneuraminic acid containing gangliosides. *Arch Biochem Biophys* 294: 427-433.
19. van Crujisen H, Gallegos Ruiz M, Van der Valk P, D de Gruijil T, Giaccone G (2009) Tissue micro array analysis of ganglioside N-glycolyl GM3 expression and signal transducer and activator of transcription (STAT)-3 activation in relation to dendritic cell infiltration and microvessel density in non-small cell lung cancer. *BMC Cancer* 9: 180.
20. Kregel U, Olsson LL, Martinez C, Talavera A, Rojas G, et al. (2004) Structure and molecular interactions of a unique antitumor antibody specific for N-Glycolyl GM3. *J Biol Chem* 279: 5597-5603.

21. Oliva JP, Valdes Z, Casaco A, Pimentel G, Gonzalez J, et al. (2006) Clinical evidences of GM3 (NeuGc) ganglioside expression in human breast cancer using the 14F7 monoclonal antibody labelled with (99m)tc. *Breast Cancer Res Treat* 96: 115-121.
22. Noguchi M, Suzuki T, Kabayama K, Takahashi H, Chiba H, et al. (2007) GM3 synthase gene is a novel biomarker for histological classification and drug sensitivity against epidermal growth factor receptor tyrosine kinase inhibitors in non-small cell lung cancer. *Cancer Sci* 98: 1625-1632.
23. Kawamura S, Ohyama C, Watanabe R, Satoh M, Saito S, et al. (2001) Glycolipid composition in bladder tumor: a crucial role of GM3 ganglioside in tumor invasion. *Int J Cancer* 94: 343-347.
24. Bousquet PA, Sandvik JA, Jeppesen Edin NF, Ute Krengel U (2018) Hypothesis: Hypoxia induces de novo synthesis of NeuGc gangliosides in humans through CMAH domain substitute. *Biochem Biophys Res Commun* 495: 1562-1566.
25. Yin J, Hashimoto A, Izawa M, Miyazaki K, Chen GY, et al. (2006) Hypoxic culture induces expression of sialin, a sialic acid transporter, and cancer-associated gangliosides containing non-human sialic acid on human cancer cells. *Cancer Res* 66: 2937-2945.
26. Labrada M, Clavell M, Bebelagua Y, de Leon J, Alonso DF, et al. (2010) Direct validation of NGcGM3 ganglioside as a new target for cancer immunotherapy. *Expert Opin Biol Ther* 10: 153-162.

Submit your manuscript at

<http://enlivenarchive.org/submit-manuscript.php>

New initiative of Enliven Archive

Apart from providing HTML, PDF versions; we also provide **video version** and deposit the videos in about 15 freely accessible social network sites that promote videos which in turn will aid in rapid circulation of articles published with us.