

# Cancer Stem Cells (Csc's): Emerging Concept and Strategy for Targeting Progression of Epithelial Ovarian Cancer

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The estimated new cases of ovarian cancer in the United States of America (USA) in 2014 is 21,980, and the estimated deaths is 14,270 [1]. It ranks 5th overall for cancer death in women, counting for 5% of all cancer deaths in women (lung, breast, colorectal, pancreatic are 1-4 respectively). At time of diagnosis, 15% of the total cases are localized, 18% are regional, and 61% are distant. In 2011, there were an estimated 188,867 women living with ovary cancer in the United States. The Standard treatment of ovarian cancer consists of surgical resection of disease followed by taxane and platinum-based chemotherapy which yields a partial response rate of about 80% and a complete response rate of 40%-60% in patients with advanced disease [1]. However, the recurrence rate is approximately 70% and five-year survival is 45% in patients with advanced disease [3-5]. It appears that the majority of ovarian cancer cells are initially chemosensitive as evidenced by the initial response rates. However, the high recurrence rates suggest development of chemoresistance. It is thought that some cells are not killed by chemotherapy, or they repopulate after exposure to chemotherapeutic agents. These cells have been described as ovarian cancer stem cells (CSCs).

The CSCs are progenitor cells [6,7], and its markers have been shown to be upregulated in cells growing in tumorspheres. In ovarian cancer, this spheroid tumor cells are thought to be related to the spread of cancer and metastasis in the peritoneal cavity. CSCs have been shown to self-renew, differentiate, and metastasize [8]. In addition, it was observed that treatment with chemotherapeutic agents results in increased drug-resistant CSCs and this leads to recurrence. Certain types of blood cancers have normal stem cells, but normal ovarian stem cell are unknown [6].

## Cancer Stem Cells Markers and its Significance

There are no specific ovarian CSCs markers and researchers rely on protein markers identified from other malignancies. These some of these proteins include CD44, CD133, CD117, ALDH1A1, and EpCAM. CD44 is a receptor that is involved in cell-cell and cell-matrix interactions and affects cellular growth, differentiation, and motility. It has been shown that CD44+/CD117+ cells had increased chemoresistance to taxane and platinum-based chemotherapy, and the ability to self-propagate [9]. It was noted that CD44+ cells were enriched in ovarian cancer patient ascites and once isolated and xenografted gave rise to tumor with both CD44+ and CD44- cells [10]. There are several strategies to target the CD44 receptor, which include binding to hyaluronic acid and osteopontin, a protein over-expressed in ovarian cancer, and contribute to receptor tyrosine kinase activation [11]. CD133 is a trans-membrane glycoprotein, and was shown that the amount of CD133 positive cells was higher in ovarian carcinoma than in normal ovarian tissue [12]. The ability of CD133+ cancer cells to generate both CD133+ and CD133- cells has been reported [13]. In addition, CD133 observed to be involved in increased tumor formation, increased chemoresistance, and the ability to recapitulate the original heterogeneous tumor [14]. CD117, also known as stem cell growth factor receptor. It has been shown to be involved in apoptosis, cell differentiation, proliferation, and cell adhesion [15]. CD117 was observed to have high expression in ovarian cancer cells [16], and cells expressing CD117 are highly tumorigenic in mouse models [17]. The Wnt/ $\beta$ -catenin pathway plays a role in the development of chemoresistance is activated by CD117 [18].

ALDH1A1 contains 19 enzymes that function as cell protectors from carcinogenic aldehydes [19]. Landen et al. [20] showed its association with chemoresistance in ovarian carcinoma. Cells that are double positive for CD133 and ALDH1A1 have a great ability to develop tumors in mouse models [21]. It has been observed that metformin decreased the population of ALDH+ cells in ovarian cancer cell lines and decreased the formation of tumor spheres in patient tumors [22]. EpCAM (CD326) is a transmembrane glycoprotein, and has been shown to have oncogenic signaling properties. Higher expression of EpCAM has been observed in metastatic ovarian tumors [23], and it is involved in epithelial to mesenchymal transition leading to metastasis [24]. CD24 is a cell membrane glycoprotein, and it was noted that the movement of CD24 from the cell membrane to the cytoplasm in borderline ovarian tumors was associated with micro-invasion and omental implants, and shorter survival time in adenocarcinoma of the ovary [25]. Moulla et al. [26], showed that the transition from membrane to cytoplasmic CD24 expression was associated with a more aggressive phenotype in borderline tumors.

### Therapeutic Approaches of Ovarian CSCs

The elimination of ovarian CSCs has been challenging in part due to heterogeneity, so the efficacy of any single drug is limited. Combined treatments that target CSCs will be a novel direction in the future; however drug treatment for CSCs may increase the risk of toxicity, as CSCs share common features with normal stem cells. The therapeutic strategies in ovarian CSCs include the following

#### 1. Nonsurface Markers

Cell surface markers (CD 133, CD 44, CD 24, and CD 117) have been used to isolate putative CSCs. The development of antibody-drug conjugates has recently achieved marked success [27]. The development of specific therapies that target biomarkers of ovarian CSCs may improve oncogenic outcome and patient's survival [28]. EpCAM is highly expressed in different tumor types, including colon, lung, pancreas, breast, head and neck, and ovary [29]. Down regulation of EpCAM may cause loss of cell-cell adhesion and promote epithelial mesenchymal transition (EMT) [30]. Catumaxomab, a monoclonal antibody against EpCAM is a trifunctional antibody, which can bind three different cell types, including tumor cells, T cells, and accessory cells (dendritic cell, macrophages, and natural killer cells) [31]. It has been used in phase III clinical trials in patients with malignant ascites [32]. In addition, two stem cell markers, Lin28 and Oct4, could serve as targeted therapy due to their roles in the maintenance of pluripotency in ovarian cancer [33]. Moreover, over expression of the Müllerian inhibiting substance (MIS) type II receptor has been reported in ovarian cancer cell lines [34]. MIS observed to inhibit the cell population with stem-like characteristics in ovarian cancer cell lines [35].

#### 2. Differentiation of Ovarian CSCs

To eliminate CSCs is to induce their differentiation, resulting in loss of their stemness property [36]. One must emphasize that, the understanding of regulation of differentiation processes is essential for designing new agents to eliminate CSCs. Yin et al. observed that TWIST-1 played a marked role in triggering differentiation of epithelial ovarian cancer (EOC) [37]. It was reported that p53 can activate two miRNAs (miR-34a and miR-145). These miRNAs shown to prompt differentiation of human embryonic stem cells [38]. It has been suggested that miRNAs may be a therapeutic target for cancer treatment [39]. In addition, retinoic acid, and its analogs are the most common differentiation agents. The all-trans-retinoic acid (ATRA)

can inhibit the proliferation and induce the differentiation via inhibition of Wnt/ $\beta$ -catenin pathway in head and neck squamous carcinoma CSC [40]. The clinical study of ATRA has shown an increased survival rate of patients with acute promyelocytic leukemia, but successful cases are limited in solid tumors [41]. Reduction of the growth of ovarian CSC via Carboplatin combined with three novel retinoid compounds was observed [42]. Moreover, linoleic acids can trigger adipocyte-like differentiation in many types of cancer cells, including ovarian cancer cell line SKOV3 [43].

#### 3. Niches of CSCs

Niches are microenvironments where CSCs reside, containing cell-cell, cell-extracellular matrix, and soluble factors that support the growth, progression, and metastasis of CSCs [44]. Bone-marrow-derived mesenchymal stem cells (MSCs) and derived cell types have been shown to secrete prostaglandin E2 and release various cytokines, which is important for the formation and progression of a tumor [45]. In addition, MSC affected metastatic ability and chemoresistance in two ovarian cancer cell lines: OVCAR3 and SKOV3 [46]. It was observed that tumorigenic ability of ovarian tumor cells was dependent on niches derived from human embryonic stem cells [47]. The hypoxic niches were beneficial for acquisition of stem-like properties of ovarian cancer cells [48].

#### 4. MicroRNAs (miRNAs)

They regulate gene expression at posttranscriptional level. Thus, miRNAs are involved in diverse biological processes, such as development and tumorigenesis [49]. The expression profile of miRNAs noted to be different between normal stem cells and CSCs (50, 51). MiR-214 was demonstrated to have the property of self-renewal and chemoresistance in ovarian CSCs [52]. In addition, miR-199a prevented tumorigenesis in xenograft model [53]. Moreover, the expression of miR-200a could reduce migrating ability of CD133+ ovarian CSCs. This was due to the inhibition of E-cadherin and ZEB2 [54]. To conclude, understanding the roles of CSCs in cancer therapy may improve the survival rate of ovarian cancer patients. It is worth-noting that, it would be difficult to effectively treat all advanced ovarian cancer patients. As, ovarian CSCs are heterogeneous, which leads to different sensitivities to the therapy used. It is reasonable to suggest, that combination therapy is the major direction for ovarian cancer treatment. The precision medicine dependent on different genomic characteristics of patients ought to be a more effective therapeutic method. In addition, the current advances in technology, such as next-generation DNA sequencing and mass spectrometry- (MS-) based proteomics, would enhance the development and implementation of ovarian cancer precision medicine.

#### References

1. American Cancer Society (2014) Cancer Facts and Figures 2014. Atlanta, GA: American Cancer Society.
2. Hoskins WPCYR, Baraket R, Markman M, Randall M (2005) Principles and practice of gynecologic oncology. 4th ed. Philadelphia: Lippincott Williams & Wilkins.
3. Leitao MM Jr, Chi DS (2009) Surgical management of recurrent ovarian cancer. *Semin Oncol* 36: 106-111.
4. Jordan CT (2004) Cancer stem cell biology: from leukemia to solid tumors. *Curr Opin Cell Biol* 16: 708-712.
5. Hope KJ, Jin L, Dick JE (2003) Human acute myeloid leukemia stem cells. *Arch Med Res* 34: 507-514.
6. Bapat SA (2010) Human ovarian cancer stem cells. *Reproduction*. 140: 33-41.

7. Dalerba P, Clarke MF (2007) Cancer stem cells and tumor metastasis: first steps into uncharted territory. *Cell Stem Cell* 1: 241-242.
8. Kitamura H, Okudela K, Yazawa T, Sato H, Shimoyamada H (2009) Cancer stem cell: implications in cancer biology and therapy with special reference to lung cancer. *Lung Cancer* 66: 275-281.
9. Zhang S, Balch C, Chan MW, Lai HC, Matei D, et al. (2008) Identification and characterization of ovarian cancer-initiating cells from primary human tumors. *Cancer Res* 68: 4311-4320.
10. Alvero AB, Chen R, Fu HH, Montagna M, Schwartz PE, et al. (2009) Molecular phenotyping of human ovarian cancer stem cells unravels the mechanisms for repair and chemoresistance. *Cell Cycle* 8: 158-166.
11. Orian-Rousseau V (2010) CD44, a therapeutic target for metastasising tumours. *Eur J Cancer* 46: 1271-1277.
12. Ferrandina G, Bonanno G, Pierelli L, Perillo A, Procoli A, et al. (2008) Expression of CD133-1 and CD133-2 in ovarian cancer. *Int J Gynecol Cancer* 18: 506-514.
13. Baba T, Convery PA, Matsumura N, Whitaker RS, Kondoh E, et al. (2009) Epigenetic regulation of CD133 and tumorigenicity of CD133+ ovarian cancer cells. *Oncogene* 28: 209-218.
14. Curley MD, Therrien VA, Cummings CL, Sergent PA, Koulouris CR, et al. (2009) CD133 expression defines a tumor initiating cell population in primary human ovarian cancer. *Stem Cells* 27: 2875-2883.
15. Miettinen M, Lasota J (2005) Kit (cd117): A review on expression in normal and neoplastic tissues, and mutations and their clinicopathologic correlation. *Applied immunohistochemistry & molecular morphology: AIMM/official publication of the Society for Applied Immunohistochemistry. Appl Immunohistochem Mol Morphol* 13: 205-220.
16. Kusumbe AP, Mali AM, Bapat SA (2009) CD133-expressing stem cells associated with ovarian metastases establish an endothelial hierarchy and contribute to tumor vasculature. *Stem Cells*. 27: 498-508.
17. Luo L, Zeng J, Liang B, Zhao Z, Sun L, et al. (2011) Ovarian cancer cells with the CD117 phenotype are highly tumorigenic and are related to chemotherapy outcome. *Exp Mol Pathol* 91: 596-602.
18. Chau WK, Ip CK, Mak AS, Lai HC, Wong AS (2013) c-Kit mediates chemoresistance and tumor-initiating capacity of ovarian cancer cells through activation of Wnt/ $\beta$ -catenin-ATP-binding cassette G2 signaling. *Oncogene* 32: 2767-2781.
19. Marchitti SA, Brocker C, Stagos D, Vasiliou V (2008) Non-P450 aldehyde oxidizing enzymes: the aldehyde dehydrogenase superfamily. *Expert Opin Drug Metab Toxicol* 4: 697-720.
20. Landen CN, Goodman B, Katre AA, Steg AD, Nick AM, et al. (2010) Targeting aldehyde dehydrogenase cancer stem cells in ovarian cancer. *Mol Cancer Ther* 9: 3186-3199.
21. Silva IA, Bai S, McLean K, Yang K, Griffith K, et al. (2011) Aldehyde dehydrogenase in combination with CD133 defines angiogenic ovarian cancer stem cells that portend poor patient survival. *Cancer Res* 71: 3991-4001.
22. Shank JJ, Yang K, Ghannam J, Cabrera L, Johnston CJ, et al. (2012) Metformin targets ovarian cancer stem cells in vitro and in vivo. *Gynecol Oncol*. 127: 390-397.
23. Bellone S, Siegel ER, Cocco E, Cargnelutti M, Silasi DA, et al. (2009) Overexpression of epithelial cell adhesion molecule in primary, metastatic, and recurrent/chemotherapy-resistant epithelial ovarian cancer: implications for epithelial cell adhesion molecule-specific immunotherapy. *Int J Gynecol Cancer* 19: 860-866.
24. Thiery JP, Acloque H, Huang RY, Nieto MA (2009) Epithelial-mesenchymal transitions in development and disease. *Cell* 139: 871-890.
25. Choi YL, Kim SH, Shin YK, Hong YC, Lee SJ, et al. (2005) Cytoplasmic CD24 expression in advanced ovarian serous borderline tumors. *Gynecol Oncol*. 97: 379-386.
26. Moulla A, Miliaras D, Sioga A, Kaidoglou A, Economou L (2013) The immunohistochemical expression of CD24 and CD171 adhesion molecules in borderline ovarian tumors. *Pol J Pathol* 64: 180-184.
27. Scott AM, Wolchok JD, Old LJ (2012) Antibody therapy of cancer. *Nature Reviews Cancer* 12: 278-287.
28. Burgos-Ojeda D, Rueda BR, Buckanovich RJ (2012) Ovarian cancer stem cell markers: prognostic and therapeutic implications. *Cancer Lett* 322: 1-7.
29. Imrich S, Hachmeister M, Gires O (2012) EpCAM and its potential role in tumor-initiating cells. *Cell Adhesion & Migration* 6: 30-38.
30. Thiery JP, Acloque H, Huang RYJ, Nieto MA (2009) Epithelial-mesenchymal transitions in development and disease. *Cell* 139: 871-890.
31. Sebastian M, Kuemmel A, Schmidt M, Schmittel A (2009) Catumaxomab: a bispecific trifunctional antibody. *Drugs Today (Barc)* 45: 589-597.
32. Seimetz D, Lindhofer H, Bokemeyer C (2010) Development and approval of the trifunctional antibody catumaxomab (anti-EpCAM\*anti-CD3) as a targeted cancer immunotherapy. *Cancer Treat Rev* 36: 458-467.
33. Peng S, Maihle NJ, Huang Y (2010) Pluripotency factors Lin28 and Oct4 identify a sub-population of stem cell-like cells in ovarian cancer. *Oncogene* 29: 2153-2159.
34. Masiakos PT, MacLaughlin DT, Maheswaran S, Teixeira J, Fuller AF Jr, et al. (1999) Human ovarian cancer, cell lines, and primary ascites cells express the human Mullerian inhibiting substance (MIS) type II receptor, bind, and are responsive to MIS. *Clin Cancer Res* 5: 3488-3499.
35. Wei X, Dombkowski D, Meirelles K, Pieretti-Vanmarcke R, Szotek PP, et al. (2010) Müllerian inhibiting substance preferentially inhibits stem/progenitors in human ovarian cancer cell lines compared with chemotherapeutics. *Proc Natl Acad Sci U S A* 107: 18874-18879.
36. Sell S (2004) Stem cell origin of cancer and differentiation therapy. *Crit Rev Oncol Hematol* 51: 1-28.
37. Yin G, Alvero AB, Craveiro V, Holmberg JC, Fu HH, et al. (2013) Constitutive proteasomal degradation of TWIST-1 in epithelial-ovarian cancer stem cells impacts differentiation and metastatic potential. *Oncogene* 32: 39-49.
38. Jain AK, Allton K, Iacovino M, Mahen E, Milczarek RJ, et al. (2012) p53 regulates cell cycle and microRNAs to promote differentiation of human embryonic stem cells. *PLoS Biol* 10: e1001268.
39. Yu Z, Li Y, Fan H, Liu Z, Pestell RG (2012) miRNAs regulate stem cell self-renewal and differentiation. *Front Genet* 3: 191-195.
40. Lim YC, Kang HJ, Kim YS, Choi EC (2012) All-trans-retinoic acid inhibits growth of head and neck cancer stem cells by suppression of Wnt/ $\beta$ -catenin pathway. *Eur J Cancer* 48: 3310-3318.
41. Soignet SL, Benedetti F, Fleischauer A, Parker BA, Truglia JA, et al. (1998) Clinical study of 9-cis retinoic acid (LGD1057) in acute promyelocytic leukemia. *Leukemia* 12: 1518-1521.
42. Whitworth JM, Londoño-Joshi AI, Sellers JC, Oliver PJ, Muccio DD, et al. (2012) The impact of novel retinoids in combination with platinum chemotherapy on ovarian cancer stem cells. *Gynecol Oncol* 125: 226-230.
43. Ruiz-Vela A, Aguilar-Gallardo C, Martinez-Arroyo AM, Soriano-Navarro M, Ruiz V, et al. (2011) Specific unsaturated fatty acids enforce the transdifferentiation of human cancer cells toward adipocyte-like cells. *Stem Cell Rev* 7: 898-909.

44. Ruiz-Vela A, Aguilar-Gallardo C, Simón C (2010) Building a framework for embryonic microenvironments and cancer stem cells. *Stem Cell Rev* 5: 319-327.
45. Li HJ, Reinhardt F, Herschman HR, Weinberg RA (2012) Cancer-stimulated mesenchymal stem cells create a carcinoma stem cell niche via prostaglandin E2 signaling. *Cancer Discov* 2: 840-855.
46. Lis R, Touboul C, Raynaud CM, Malek JA, Suhre K, et al. (2012) Mesenchymal cell interaction with ovarian cancer cells triggers pro-metastatic properties. *PLoS One* 7: e38340.
47. Katz E, Skorecki K, Tzukerman M (2009) Niche-dependent tumorigenic capacity of malignant ovarian ascites-derived cancer cell subpopulations. *Clin Cancer Res* 15: 70-80.
48. Liang D, Ma Y, Liu J, Trope CG, Holm R, et al. (2012) The hypoxic microenvironment upgrades stem-like properties of ovarian cancer cells. *BMC Cancer* 12: 201-211.
49. Bartel DP (2009) MicroRNAs: target recognition and regulatory functions. *Cell* 136: 215-233.
50. Lavon I, Zrihan D, Granit A, Einstein O, Fainstein N, et al. (2010) Gliomas display a microRNA expression profile reminiscent of neural precursor cells. *Neuro Oncol* 12: 422-433.
51. van Jaarsveld MTM, Helleman J, Berns EMJJ, Wiemer EA (2010) MicroRNAs in ovarian cancer biology and therapy resistance. *Int J Biochem Cell Biol* 42: 1282-1290.
52. Xu CX, Xu M, Tan L, Yang H, Permeth-Wey J, et al. (2012) MicroRNA MiR-214 regulates ovarian cancer cell stemness by targeting p53/Nanog. *J Biol Chem* 287: 34970-34978.
53. Cheng W, Liu T, Wan X, Gao Y, Wang H (2012) MicroRNA-199a targets CD44 to suppress the tumorigenicity and multidrug resistance of ovarian cancer-initiating cells. *FEBS J* 279: 2047-2059.
54. Wu Q, Guo R, Lin M, Zhou B, Wang Y (2011) MicroRNA-200a inhibits CD133/1+ ovarian cancer stem cells migration and invasion by targeting E-cadherin repressor ZEB2. *Gynecol Oncol* 122: 149-154.

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